PRO-MEASURE™ Protein Measurement Solution

Cat. No.

21011

100 ml

DESCRIPTION

Protein quantification is one of the important steps in protein research such as W estern blot assay. Protein assay methods are diverse, and there is one which abs orbs at 280nm without special reagents or operation. This method is simple, yet i t also has a shortcoming not being able to analyze protein assay when no amino acid such as phenylalanine, tryptophan or tyrosine are present. Neither will it be used commonly due to its DNA absorbance interference (AI). The other methods are Lowry assay or BCA assay, which are highly sensitive for protein quantificatio n, but is guite inconvenient and time-consuming. Contrarily, Bradford method is t he most commonly used one (within 10min), which can detect the minimum amo unt of protein and is very $% \left(1\right) =\left(1\right) =\left(1\right) =\left(1\right)$ where $\left(1\right) =\left(1\right)$ nient than Bradford assay, declining AI of solution itself so that the background a bsorbance is maintained very low.

CHARACTERISTIC

- · Very simple usage steps
- · Short measurement time within 10min.
- High sensitivity to detect 10-25 g/ml protein.
- · Lower background interference.
- · Able to to quantify protein by minimum amount of reagents.

CONTENTS

- PRO-MEASURE™ solution(10x)

100 ml

· Standard solution (BSA, 1 mg/ml)

1 ml

STORAGE

Store at 4°C, and then stable for more than 1 year.

PREPARATION BEFORE USE

Before use, dilute 1 part PRO-MEASURE™ solution by adding 9 parts of sterile wat er. For example, if you are to produce 50 ml dilution solution, mix 5 ml PRO-MEAS URE™ with 45 ml sterile water.

CONSIDERATION BEFORE USE

1. In protein quantification, there can be an error due to high AI, depending on the solution type used for protein assay. Therefore, one must perform calibration u sing a blank (background absorbance) to nullify possible errors. For example, it has to be handed carefully if using a solution containing detergents such as Tri ton X-100, NP-40, deoxycholate etc., for they effect the absorbance of protein a ssay. iNtRON PRO-PREP™ (Cat. No.17081) can extract protein without affectin g the protein assay.

- 2. In protein assay, one must use the original solution for dilution. To keep an err or to minimum, one can perform dilution by using the protein extracting solutio n or using sterile water if necessary.
- 3. When measuring absorbance, use plastic cuvette. Because, this solution can c oat the surface of glass cuvette. As the results, this solution give effect for abs orbance measurement. PRO-MEASURE™ solution on glass cuvette surface can be removed by MtOH washing.
- 4. When quantification of protein, normally assay in duplicate or triplicates. Beca use we can decrease an measurement error.
- 5. For each protein measurement, one should calculate standard curve. By the wa y, the equation of iNtRON is suitable for any experiment with relative equal volu me (Western blot, etc).
- 6. When measuring absorbance at 595nm, you can use cuvette or 96well-micro p
- 7. Suitable reaction time is within 2-10 min after the addition of samples. Absorb ance will increase over time, solution should incubate at room temperature for no more than 1 hour.

PROTOCOL

Following method is brief summary.

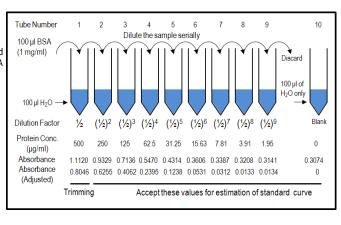
- 1. Prepare sample solution of 100 I through dilution of protein.
- Add 1 ml of diluted PRO-MEASURE™ solution.
- 3. Incubate for 2 min at RT, measure absorbance at 595 nm.
- 4. Calculate concentration of protein by formula.

STANDARD CURVE

- This experiment method is to determine standard curve for BSA (bovine serum albumin) (Figure provided).
- The standard curve presented in this manual can be varied depending on enviro nment and equipment of the actual experiments, and path-length of cuvette. Th erefore we recommend creating standard curve every experiment.
- 1. Prepare two sets of 1.5 ml tubes each set with 10 tubes, add 100µl of ste rile water.
- 2. Add 100µl of BSA solution to 1st tube and mix, 100µl of solution transfer by pip etting from this tube(1st tube) to 2nd tube(1/2 dilution). Prepare for until numb er 9 tube by 1/2 dilution method. Number 10 tube add 100µl of dH2O, apply to a s blank control.
- 3. Add 1 ml of PRO-MEASURE™ dilutes to each tubes, incubate for 2 min, measur e absorbance at 595 nm.
- 4. Calculate standard curve through OD and concentration of BSA.
- 5. Introduce to equation; Y (protein concentration) = $500 \times A_{595} 13.5$

[Step 1] Add 100µl of dH O to each tubes and dilute serially by ad dition of 100µl of BSA solution.

[Step 2] Add 1 ml of diluted $\mathsf{PRO}\text{-}\mathsf{MEASURE}^{\mathsf{TM}}$ solution, measure at 595nmafter incu bating for 2 min.



Calculate standard curve through OD₅₉₅ and concentration of BSA. 0.7 v = 0.002x + 0.023 $R^2 = 0.972$ 0.6 Absorbance at OD595nm 0.5 0.3 0.2 0.1 50 200 250 150 BSA Standard (µg/ml)

